

Biological Chemistry Laboratory
Biology 3515/Chemistry 3515
Spring 2023

Lecture 13

Determination of K_m and V_{max}

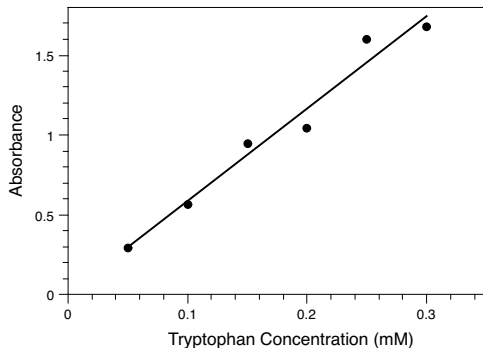
21 February 2023

©David P. Goldenberg

University of Utah

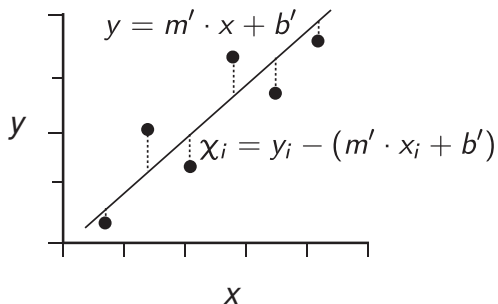
goldenberg@biology.utah.edu

From Quiz 1, Problem 3a



- Should we fit a different function to the data?
 - Is there a systematic deviation of the data from the linear relationship?
 - Does the linear relationship (or a different function) represent a good model to account for the data?

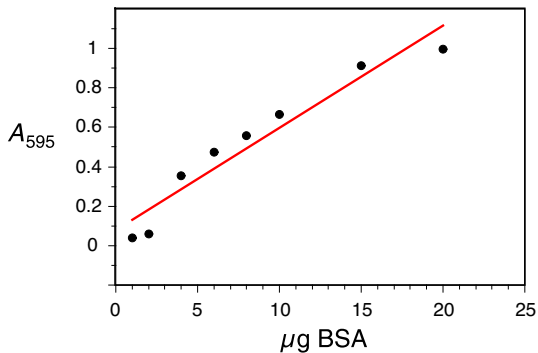
The Method of Least Squares: χ^2



$$\begin{aligned}\chi^2 &= \sum \chi_i^2 \\ &= \sum (y_i - (m' \cdot x_i + b'))^2\end{aligned}$$

- Adjust m' and b' to minimize the value of χ^2 for the particular values of x_i and y_i in the experimental data set.
- The method can be applied to other functions to fit parameters.

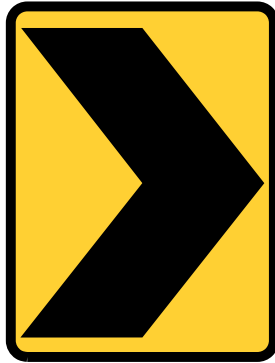
The Coefficient of Determination, R^2



$$R^2 = 0.93$$

- R^2 represents the fraction of the variation that is accounted for by the fit function.
- R^2 usually lies between 0 and 1.
- R^2 can be negative for certain functions and data sets!

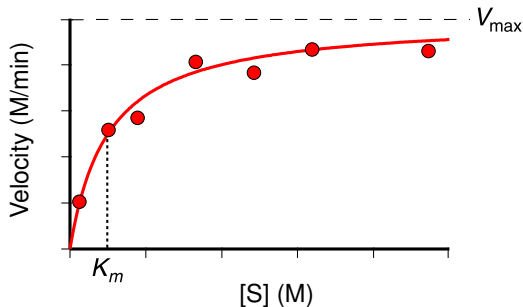
Warning!



Direction Change

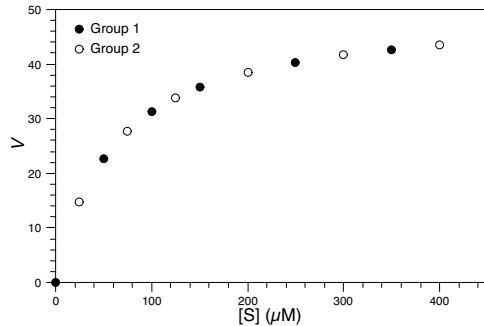
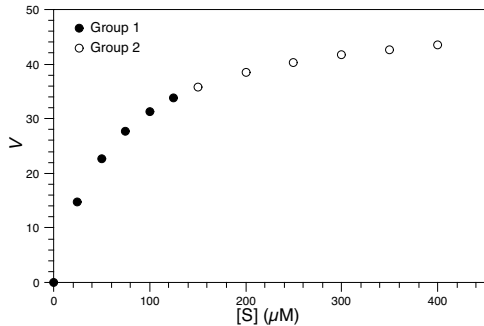
Determining K_m and V_{max}

Experiment 3, Part D: Velocity as a Function of Substrate Concentration



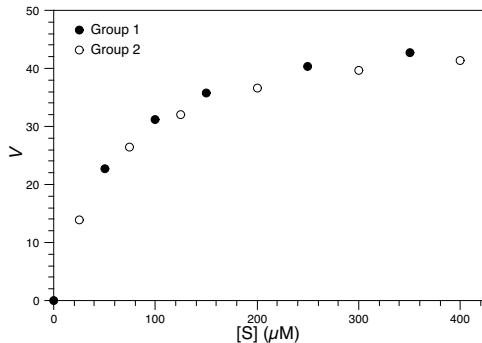
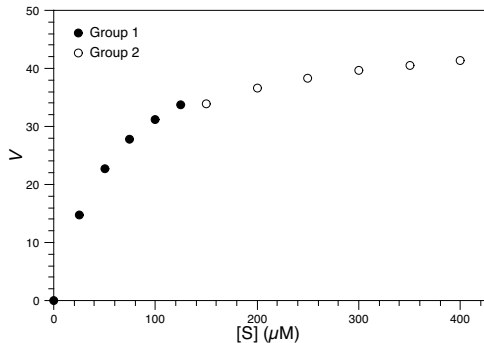
- To reliably estimate both K_m and V_{max} , substrate concentrations must cover range both below and above K_m .
- We will use eleven substrate concentrations, plus a control without substrate.
- Two groups of six reactions.
- How should the reactions be grouped?

Two Ways to Group the Reactions



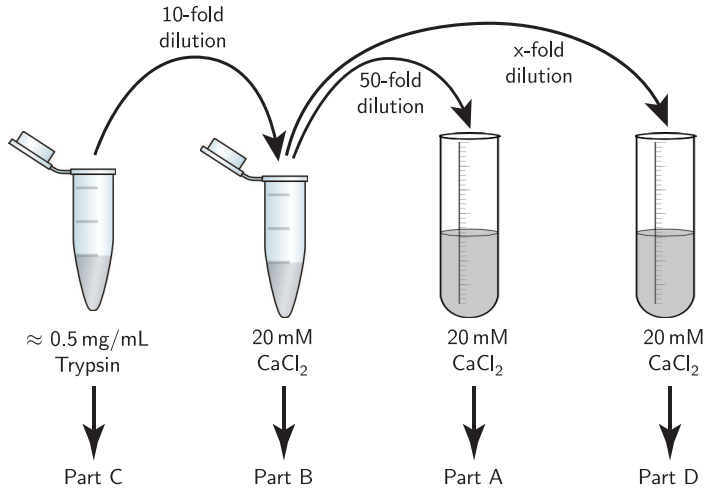
- Is one way better than the other?
- What if something changes between the reactions?

Two Ways to Group the Reactions

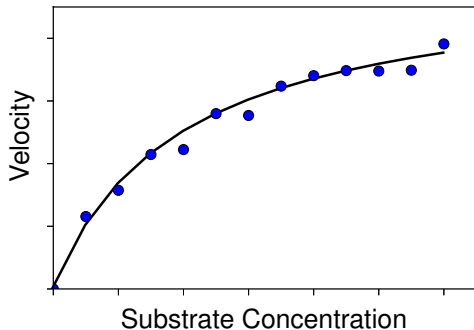


- 1–6, 7–12 grouping: changes between reaction groups may be hard to detect.
- Odd–even grouping: changes between reaction groups are easier to detect.
Fitting data together averages effects more evenly.

Dilutions of Trypsin Solutions for Experiment 3



Analysis of Data from the V versus $[S]$ Experiment



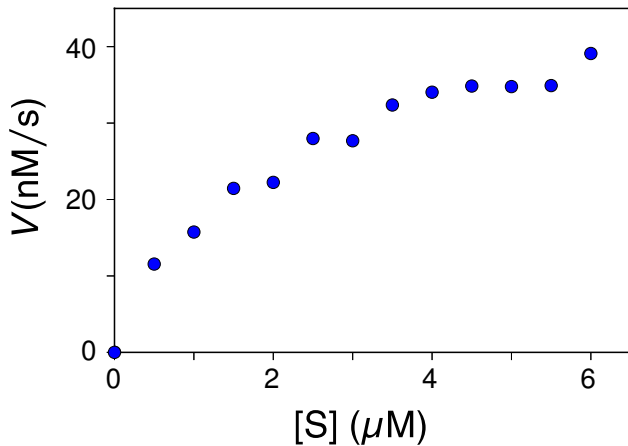
- We want to fit the experimental data to the Michaelis-Menten Equation:

$$V = \frac{[S]V_{\max}}{[S] + K_m}$$

- From the fit, we obtain estimates of K_m and V_{\max} .

Clicker Question #1

Estimate V_{\max} from the graph:



A) 30 nM/s

B) 40 nM/s *

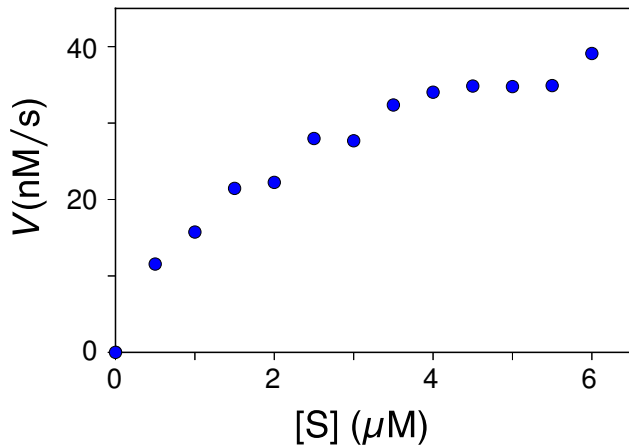
C) 50 nM/s

D) 80 nM/s

*close enough for credit.

Clicker Question #2

Estimate K_m from the graph:



- A) $1 \mu\text{M}$
- B) $2 \mu\text{M}$
- C) $5 \mu\text{M}$
- D) $10 \mu\text{M}$

A Classic Method for Analyzing Enzyme Kinetics Data

- Rearrangement of the Michaelis-Menten Equation:

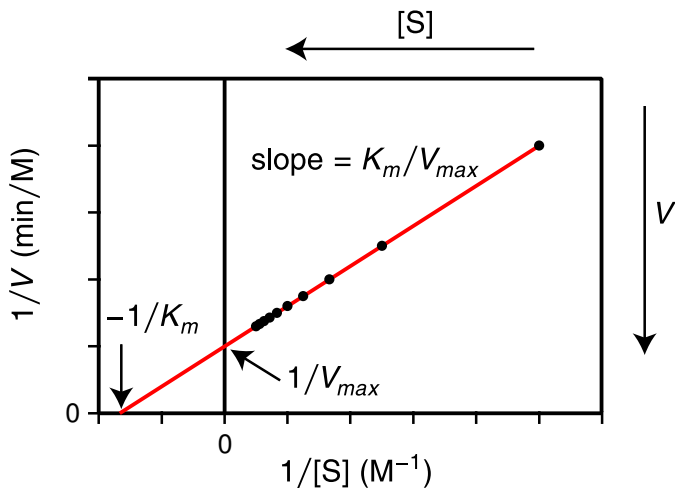
$$V = \frac{[S]V_{\max}}{[S] + K_m}$$

$$\frac{1}{V} = \frac{[S] + K_m}{[S]V_{\max}} = \frac{[S]}{[S]V_{\max}} + \frac{K_m}{[S]V_{\max}}$$

$$\frac{1}{V} = \frac{1}{[S]} \cdot \frac{K_m}{V_{\max}} + \frac{1}{V_{\max}}$$

- A “double-reciprocal” plot of $1/V$ versus $1/[S]$ should generate a straight line with a slope of K_m/V_{\max} and an intercept of $1/V_{\max}$ on the $1/V$ axis.

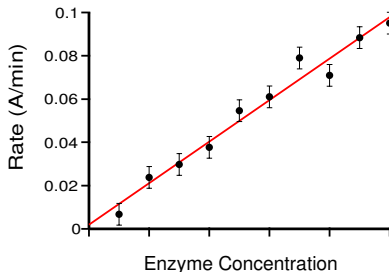
The Lineweaver-Burk Plot



- If the data are perfect, this plot gives good estimates of K_m and V_{max} .
- But, experimental error in V can lead to strange effects!

Experimental Error and Uncertainty

- Error bars for rate measurements are of approximately constant size (*e.g.*, ± 0.005 A/min), rather than a constant percentage of the measurement.



- For 0.1 A/min, ± 0.005 A/min = $\pm 5\%$.
- For 0.01 A/min, ± 0.005 A/min = $\pm 50\%$.
- Least-squares fitting works well if the *absolute* uncertainties of all data points are approximately equal.

What Happens When We Take Reciprocals?

■ $V = 0.1 \pm 0.005$

$$\frac{1}{0.105} = 9.52,$$

$$\frac{1}{0.095} = 10.5,$$

$$\frac{1}{V} = 10 \pm 0.5$$

■ $V = 0.01 \pm 0.005$

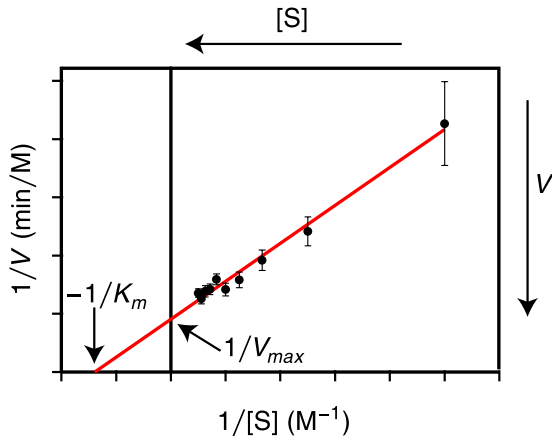
$$\frac{1}{0.015} = 66.7,$$

$$\frac{1}{0.005} = 200,$$

$$\frac{1}{V} = 100 \pm 50$$

- The values of $1/V$ derived from small velocities can have very large absolute errors.

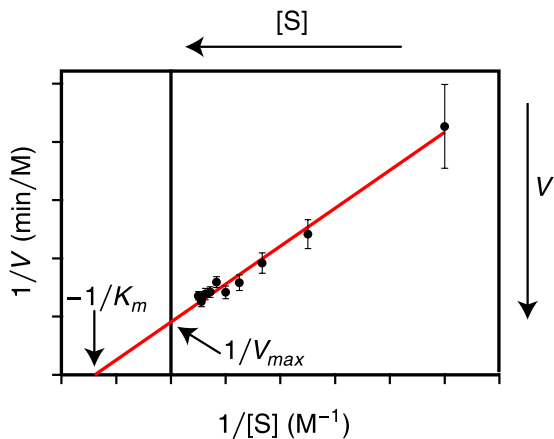
The Effects on a Lineweaver-Burk Plot



- Errors in the least precise measurements (low V) can cause large changes in the line fit to the Lineweaver-Burk plot.

Clicker Question #3

Which parameter is likely to be more sensitive to errors in a Lineweaver-Burk plot?



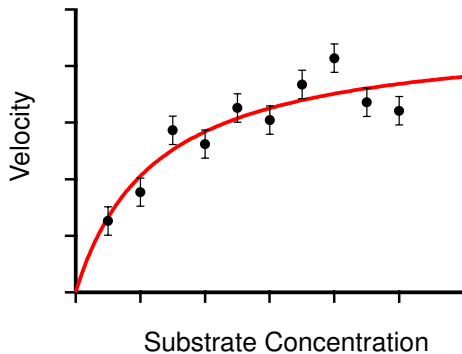
A) K_m *

B) V_{max}

* Provided that values of V approach V_{max} .

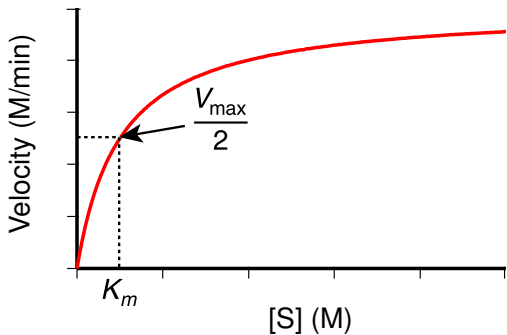
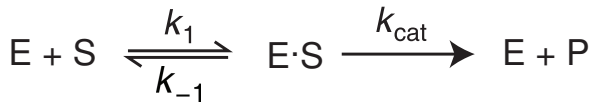
Two Ways to Deal with This Problem

- Use Lineweaver-Burk, but weight data according to uncertainties in $1/V$.
- Fit velocity data directly to the Michaelis-Menten equation using non-linear least-squares method.



- Equal errors in V are weighted equally.

Interpreting K_m



$$K_m = \frac{k_{-1} + k_{\text{cat}}}{k_1}$$

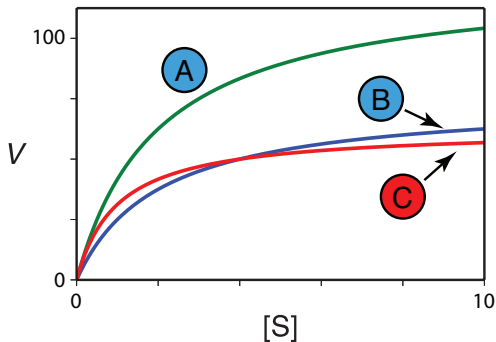
$$V = V_{\text{max}} \frac{[S]}{[S] + K_m}$$

$$V = \frac{V_{\text{max}}}{2} \quad \text{when} \quad [S] = K_m$$

- When $[S] = K_m$, half of total enzyme has substrate bound.
- The larger K_m is, the more substrate is required to reach $V_{\text{max}}/2$, or any specified fraction of V_{max} .

Clicker Question #4:

Data for three substrates with the same enzyme.



Which substrate binds most tightly to the enzyme?

No wrong answers, for now.

A Closer Look at Binding and K_m : K_m versus K_d



- K_m is defined in terms of the rate constants:

$$K_m = \frac{k_{-1} + k_{\text{cat}}}{k_1}$$

- K_d is the equilibrium constant for dissociation.

$$K_d = \frac{[E]_{\text{eq}}[S]_{\text{eq}}}{[E \cdot S]_{\text{eq}}} = \frac{k_{-1}}{k_1}$$

A large K_d indicates weak binding.

K_m versus K_d

$$K_m = \frac{k_{-1} + k_{\text{cat}}}{k_1}$$

$$K_d = \frac{[E]_{\text{eq}}[S]_{\text{eq}}}{[E \cdot S]_{\text{eq}}} = \frac{k_{-1}}{k_1}$$

- If $k_{\text{cat}} \ll k_{-1}$, the E·S complex is more likely to dissociate than undergo catalysis:

$$K_m \approx \frac{k_{-1}}{k_1} = K_d$$

- In general, $K_m \geq K_d$
- Strength of equilibrium binding may be greater than indicated by K_m .