Biological Chemistry Laboratory Biology 3515/Chemistry 3515 Spring 2023

Lecture 17:

Irreversible Serine Protease Inhibitors

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The Trypsin Active Site



Clicker Question #1

Which residue carries out nucleophilic attack on the substrate?



Clicker Question #2

Which residue deprotonates the nucleophilic oxygen atom?



Irreversible Inhibition of Trypsin by AEBSF

4-(2-aminoethyl)-benzenesulfonyl fluoride



Reaction is specific for the catalytic Ser residue.

Reaction is irreversible.

An Earlier Irreversible Inhibitor of Serine Proteases: Diisopropyl Fluorophosphate



- First synthesized in the 1930's as a potential chemical weapon with neurotoxic effects.
- Found to inhibit esterases and proteases.
- Still widely used as a pesticide.

Acetylcholine Esterase



- Acetylcholine is a major neurotransmitter in vertebrates, insects and other animals.
- Esterase reaction is very similar to peptide hydrolysis.
- Enzyme uses a catalytic triad (Ser-His-Glu).
- Enzyme is inhibited by DIFP and other serine-reactive agents.

A Cholinergic Synapse



- Irreversible inhibition of acetylcholine esterase is lethal.
- Mild reversible inhibition may be therapeutic.
- AEBSF is much less toxic than DIFP.

Two Antidotes for Organophosphate Poisoning



Atropine



- Isolated from Atropa belladonna, "deadly nightshade".
- Inhibitor of acetylcholine receptors.
- Causes many symptoms, including dilation of pupils.
- "Bella dona" is Italian for "beautiful lady". Plant extracts were used by women to dilate their eyes.
- Shorter acting drugs are used by ophthalmologists for dilation.

A Cholinergic Synapse



- Atropine competes with acetylcholine for binding to acetylcholine receptors.
- Atropine is an acetylcholine antagonist; binds to receptor without activating.
- Blocks synaptic transmission; better than continuous activation.

Pralidoxime



- Reactivates inhibited acetylcholine esterase.
- What makes this molecule special?
- Hydroxyl has unusually low pK_a , ≈ 8 .
- Oxygen is especially reactive.

Inactivation and Reactivation of Acetylcholine Esterase



Mercey, G., Verdelet, T., Renaou, J., Kiliachynai, M., Baatli, R., Nachon, F., Jean, L. & Renard, P.-Y. (2012). Reactivators of acetylcholinesterase inhibited by organophosphorous nerve agents. *Acc. Chem. Res.*, 45, 756–766. http://dx.doi.org/10.1021/ar2002864

Organophosphorous Poisoning and Treatment

- ~200,000 deaths per year by self-administered organophosphorous pesticides in rural Asia.
- Muscineric acetylcholine antagonists (*e.g.*, atropine) are generally the initial treatment.
- Oxime reactivators (*e.g.*, pralidoxime) are often used in conjunction with atropine.
 - Oximes are not very general. Different oximes are specific for different organophosphorous toxins.
 - Efficacy of oximes is debated.

Eddleston, M., Buckley, N. A., Eyer, P. & Dawson, A. H. (2008). Management of acute organophosphorous pesticide poisoning. *The Lancet*, 371, 16–22. https://doi.org/10.1016/S0140-6736(07)61202-1

4-(2-aminoethyl)-benzenesulfonyl fluoride



Experimental Protocol for Studying Irreversible Inhibition

 Follow the reaction by measuring enzymatic activity at increasing times after mixing enzyme and inhibitor.



- For each sample withdrawn, measure reaction velocity.
- $V \propto$ concentration of uninhibited enzyme.
- Time for assay must be short relative to time of inactivation.

Clicker Question #3

How does the concentration of active enzyme change with time?



All answers count for now.

Kinetics of Irreversible Inactivation

- $\blacksquare \mathsf{E} + \mathsf{I} \to \mathsf{E} \mathsf{-} \mathsf{I}$
- Second-order kinetics:

$$\frac{d[\mathsf{E}]}{dt} = \frac{d[\mathsf{I}]}{dt} = -k_2[\mathsf{I}][\mathsf{E}]$$

If initial concentrations of enzyme and inhibitor are equal:



This is not an exponential decay function!

Both [I] and [E] decrease with time, and both decreases contribute to reduced rate with time.

Pseudo First-Order Kinetics

 \blacksquare If [I] \gg [E], [I] will remain approximately constant during the reaction.

$$\frac{d[\mathsf{E}]}{dt} = -\underbrace{k_2[\mathsf{I}]}_{\text{constant}} \cdot [\mathsf{E}]$$

• Define a pseudo first-order rate constant: $k_{app} = k_2[I]$

$$\frac{d[\mathsf{E}]}{dt} = -k_{\mathsf{app}}[\mathsf{E}]$$

Rearrange and integrate the rate expression:

$$\int_{[\mathsf{E}]_0}^{[\mathsf{E}]} \frac{d[\mathsf{E}]}{[\mathsf{E}]} = \int_{t=0}^t -k_{\mathsf{app}} dt$$

 $[E]_0 =$ Initial enzyme concentration.

Pseudo First-Order Kinetics

Integrated rate expression:

$$\ln\left(\frac{[\mathsf{E}]}{[\mathsf{E}]_0}\right) = -k_{\mathsf{app}}t, \qquad \frac{[\mathsf{E}]}{[\mathsf{E}]_0} = e^{-k_{\mathsf{app}}t}$$

Standard plot:



Semi-logarithmic plot:



The exponential function, $y = e^x$, is its own derivative:

$$\frac{de^{x}}{dx} = e^{x}$$

■ More generally, if *k* is a constant:

$$\frac{de^{kx}}{dx} = ke^{kx}$$

- Why is this important?
- There are many physical and biological processes for which the rate of change is proportional to the quantity that is changing!

Clicker Question #4 (Multiple answers possible!)

Which of these represents an exponential function: $y = e^{kx}$?



Exponential Growth



Example: Growth of bacteria:

- Each bacterium has an equal probability of dividing during a given time period.
- The number of new bacteria in a short time period is proportional to the number already present.

Exponential Decay



Example: Radioactive decay:

- Each nucleus has an equal probability of decaying during a given time period.
- The number of nuclei that decay in a short time period is proportional to the number of nuclei present.

Back to Pseudo First-Order Kinetics

Second-order kinetics:

$$\frac{d[\mathsf{E}]}{dt} = \frac{d[\mathsf{I}]}{dt} = -k_2[\mathsf{I}][\mathsf{E}]$$

■ If [I] does not change significantly, we can define an apparent rate constant:

$$k_{\mathsf{app}} = k_2[\mathrm{I}]$$

Pseudo first-order kinetics

$$\frac{d[\mathsf{E}]}{dt} = -k_{\mathsf{app}}[\mathsf{E}]$$

Integrated rate expression:

$$\ln\left(\frac{[\mathsf{E}]}{[\mathsf{E}]_0}\right) = -k_{\mathsf{app}}t, \qquad \qquad \frac{[\mathsf{E}]}{[\mathsf{E}]_0} = e^{-k_{\mathsf{app}}t}$$

Data Interpretation

Estimate k_{app} from fit of $[E]/[E]_0$ versus time.

Calculate second-order rate constant, k_2 from k_{app}

$$k_2 = k_{\rm app}/[{
m I}]$$

Can use estimate of k₂ to predict kinetics of inactivation at other inhibitor concentrations.