

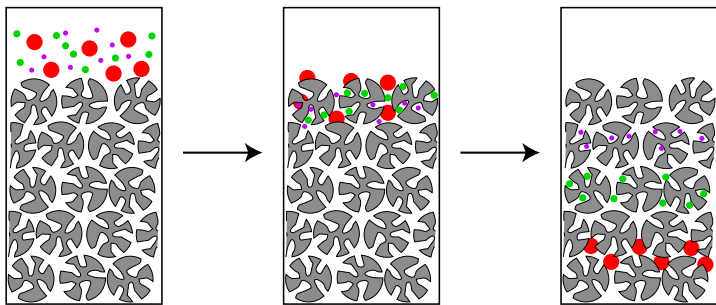
Biological Chemistry Laboratory
Biology 3515/Chemistry 3515
Spring 2023

Lecture 26:

More on Gel Filtration Chromatography
and
Experiment 6

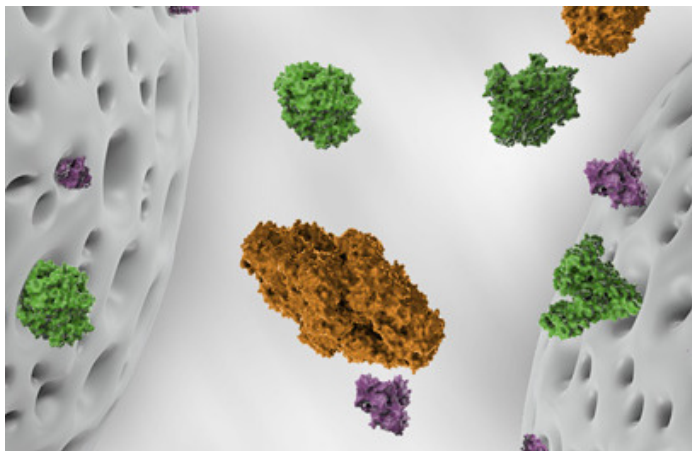
13 April 2023
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Gel Filtration Chromatography



- Partition between stationary and mobile phases depends on ability of molecules to enter pores of the beads.
- K_{av} is the fraction of the bead volume that a molecule can enter.

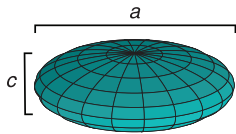
How Does Shape Affect K_{ave} (or Elution Volume)?



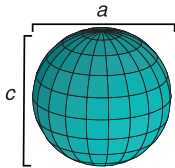
Cartoon by GE HealthCare:

<http://www.gelifesciences.com/webapp/wcs/stores/servlet/catalog/en/GELifeSciences-us/brands/superdex/>

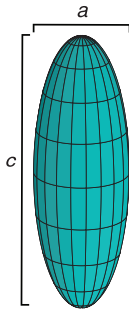
Useful Approximations for Non-spherical Proteins



Oblate ellipsoid
(M&M)
 $c/a < 1$



Sphere
 $c/a = 1$



Prolate ellipsoid
(cigar)
 $c/a > 1$

- Drawings scaled to represent equal volumes.
- Axial ratio, c/a , defines asymmetry.
- For a given volume (\propto MW) which is more likely to enter a pore?

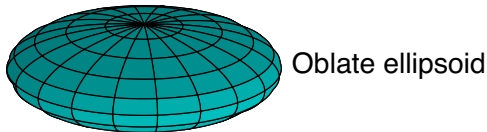
Clicker Question #1

If the volumes are the same, which will elute first from a gel filtration column?

A) The sphere.



B) The oblate ellipsoid.



C) They will elute together.

All answers count for now.

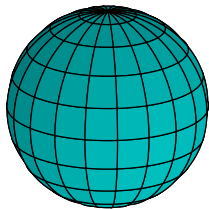
Clicker Question #2

If the volumes are the same, which will elute first from a gel filtration column?

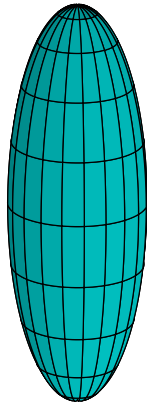
A) The sphere.

B) The prolate ellipsoid.

C) They will elute together.



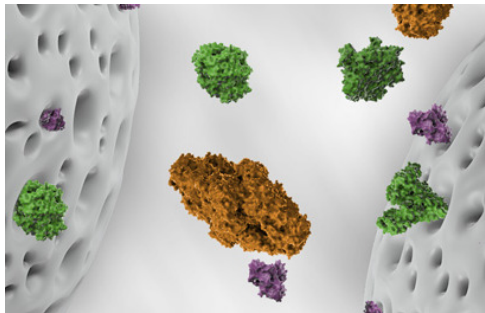
Sphere



Prolate ellipsoid

All answers count for now.

Why Does a Prolate Ellipsoid Elute First?



- The prolate ellipsoid should be able to enter smaller pores than a sphere of equal volume.
- But, ellipsoid has to be in the right orientation to enter the smaller pore.
- Most orientations make it *harder* to enter a smaller pore.
- In thermodynamic terms, the ellipsoid has to lose entropy to enter the stationary phase.
- Elution volume reflects equilibrium between stationary and mobile phase.

Effects of Size and Shape on K_{ave} (or Elution Volume)

- For molecules with the same overall shape, K_{ave} and elution volume decrease with increased molecular weight (over suitable range for a given media).
- For molecules with different shapes, elution volumes may not reflect molecular weights.
 - When compared to \approx spherical proteins, more asymmetric molecules will appear to have larger molecular weights than they really do.
 - For molecules with a range of shapes, K_{ave} is best correlated with their frictional coefficients and diffusion rates in solution.
 - “Stokes radius” or “hydrodynamic radius”, radius of a smooth sphere with the same frictional coefficient.
Can be combined with other information (such as sedimentation velocity) to better determine molecular weight and shape.

Outline of Chromatography Experiment

Day 1:

1. Prepare column

- Pour hydrated beads into glass column.
- Flow several column-volumes of buffer through column.

2. Calibrate column

- Apply a mixture of a large and a small molecule, blue dextran ($MW \approx 2,000,000$) and phenol red ($MW\ 354$).
- Elute column and collect fractions.
- Measure absorbance of fractions.

3. Record image of SDS gel.

Day 2:

1. Separate trypsin and benzamidine

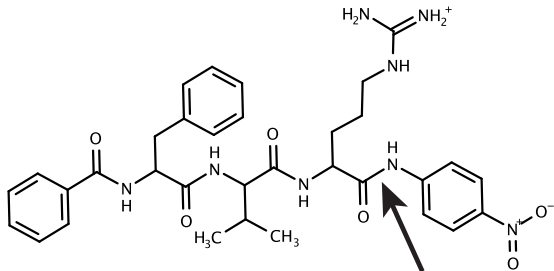
- Mix trypsin and benzamidine - inhibit trypsin activity.
- Apply mixture to column.
- Elute with buffer.
- Measure A_{280} of fractions.
- Measure trypsin activity of peak fraction. Is activity restored?

The Last Experiment: Some Considerations

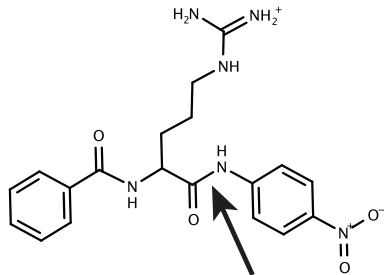
- Compare column sample with inhibited and uninhibited control samples.
- To make a meaningful comparison, the trypsin concentrations must be the same in all of the assays.
- To be detected in the column fractions, the trypsin concentration must be relatively high. ≈ 0.1 mg/mL.
- By comparison, the trypsin samples used in our previous rate measurements had concentrations of ≈ 0.001 mg/mL.
- If we dilute the trypsin samples in this experiment to make the rate slow enough to measure, we will also dilute any inhibitor present.
- The solution: Use a substrate that is cleaved more slowly than Bz-Phe-Val-Arg-*p*-NA.

Two Chromogenic Substrates for Trypsin

Bz-Phe-Val-Arg-*p*-nitroanilide

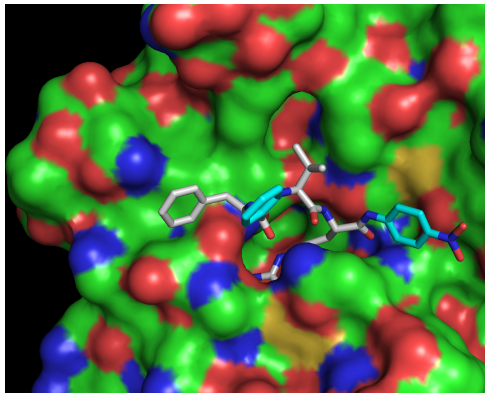


Bz-Arg-*p*-nitroanilide

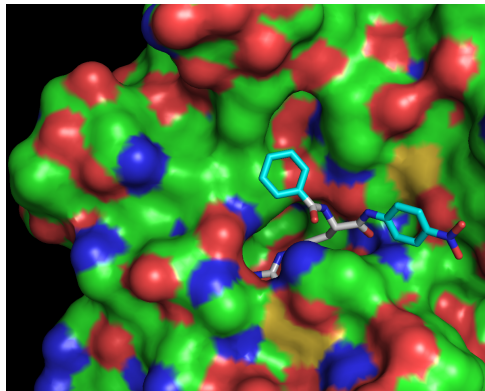


Models of the Two Substrates Bound to Trypsin

Bz-Phe-Val-Arg-*p*-nitroanilide



Bz-Arg-*p*-nitroanilide



- How will K_m and k_{cat} for the two substrates differ?

Clicker Question #3

For Bz-Arg-*p*-nitroanilide, compared to Bz-Phe-Val-Arg-*p*-nitroanilide,

- A) K_m will be greater, and k_{cat} will be the same.
- B) k_{cat} will be smaller, and K_m will be the same.
- C) K_m will be greater, and k_{cat} will be smaller.
- D) K_m will be greater, and k_{cat} will be greater.
- E) K_m will be smaller, and k_{cat} will be smaller.

All answers count for now.

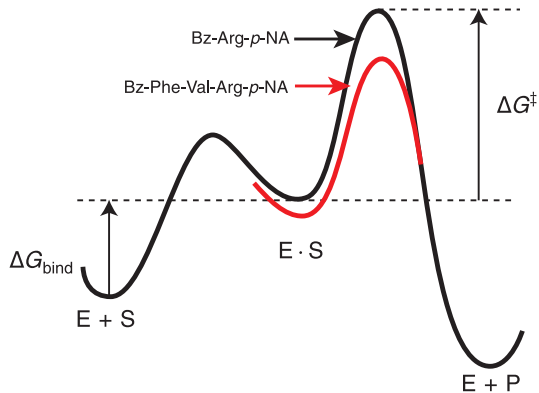
Kinetic Parameters for the Two Substrates

	K_m	k_{cat}	k_{cat}/K_m
Bz-Arg- <i>p</i> -nitroanilide	600 μM	1 s^{-1}	$1.7 \times 10^3 \text{ s}^{-1} \text{ M}^{-1}$
Bz-Phe-Val-Arg- <i>p</i> -nitroanilide	60 μM	10 s^{-1}	$1.7 \times 10^5 \text{ s}^{-1} \text{ M}^{-1}$

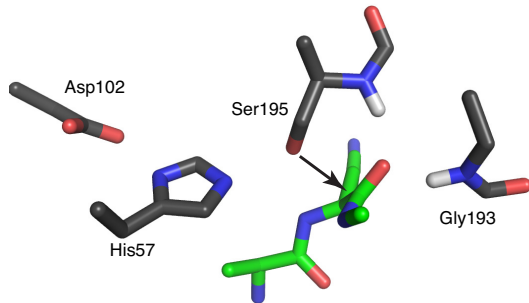
Adding Phe and Val to Bz-Arg-*p*-nitroanilide

- Decreases K_m by ≈ 10 - fold.
- Increases k_{cat} by ≈ 10 -fold.
- Increases k_{cat}/K_m by ≈ 100 -fold.

Why Does Making the Substrate Bigger Increase k_{cat} ?

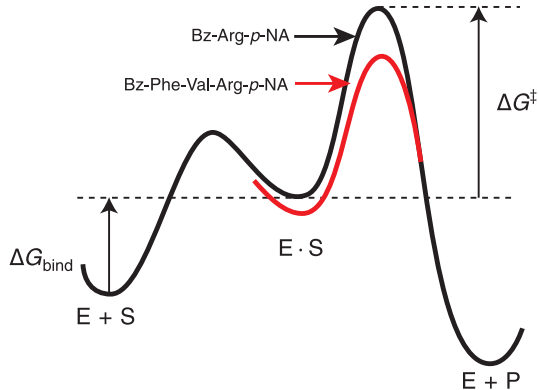


Enzyme-Substrate Complex

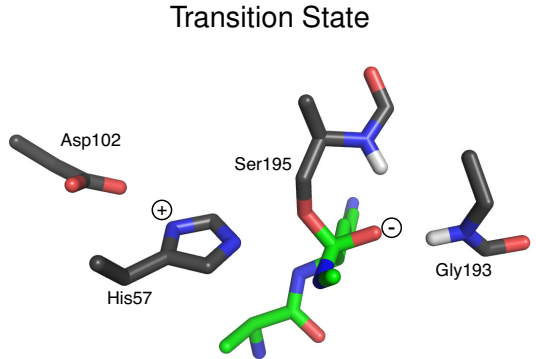


- Extra atoms of the larger substrate stabilize the $E \cdot S$ complex, relative to the free enzyme and substrate.
- But, the extra atoms stabilize the transition state *more*.

Why Does Making the Substrate Bigger Increase k_{cat} ?



- Extra atoms of the larger substrate stabilize the $E \cdot S$ complex, relative to the free enzyme and substrate.
- But, the extra atoms stabilize the transition state *more*.



- A hypothesis: The extra interactions with Phe and Val in Bz-Phe-Val-Arg-*p*-NA push the carbonyl group towards the tetrahedral conformation of the transition state.